# ORIGINAL ARTICLE

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# In vitro phase II comparison of the cytotoxicity of a novel platinum analog, nedaplatin (254-S), with that of cisplatin and carboplatin against fresh, human ovarian cancers

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**Abstract** *Purpose*: To compare the in vitro cytotoxicity of nedaplatin, an investigational platinum analog, with that of the standard platinum agents, cisplatin and carboplatin, against fresh human, epithelial ovarian cancers. Methods: The Hamburger-Salmon human tumor colony-forming assay (HTCA) was used to measure the chemosensitivity of 36 fresh tumor samples obtained during initial exploratory laparotomy from patients with newly diagnosed stage III-IV epithelial ovarian cancer who had received no prior chemotherapy or radiation therapy. Tumor samples were exposed to the platinum analogs for 1 h at concentrations of 10 and 100 µg/ml of nedaplatin and cisplatin and 100 and 1000 μg/ml of carboplatin. The resulting survival data were used to estimate the IC<sub>50</sub> (drug concentration associated with 50% inhibition of tumor colony forming units, TCFUs) of each of the platinum analogs for each of the tumor samples, as well as the estimated survival following exposure to clinically achievable drug levels (i.e. the ultrafiltrable platinum area under the plasma disappearance curve, AUC, achieved in cancer patients following administration of standard or phase II doses). Results: At the lowest concentration tested (i.e. 10 µg/ml nedaplatin and cisplatin and 100 µg/ml carboplatin) the percentages of tumor samples which were sensitive (as defined by 50% or less survival of TCFUs as compared with controls) were 42, 50, and 40% for nedaplatin, cisplatin and carboplatin, respectively. The median IC50 values were 28.5, 12 and

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121 µg/ml for nedaplatin, cisplatin and carboplatin, respectively. The estimated percentage of tumors sensitive to clinically achievable dose levels was 42% for nedaplatin and 36% for cisplatin and carboplatin. Nedaplatin and carboplatin proved relatively crossresistant with cisplatin in vitro; of the 18 tumor samples which were resistant to cisplatin, only 5 (28%) were sensitive to nedaplatin and 3 of 17 (18%) were sensitive to carboplatin. Conclusion: Nedaplatin was associated with cytotoxicity similar to cisplatin and carboplatin in this study. Although nedaplatin appears to be crossresistant with cisplatin, its high rate of in vitro cytotoxicity, relative lack of neurotoxicity and nephrotoxicity, and large in vivo bioavailability establish nedaplatin as a promising platinum analog for further clinical development as a salvage and primary chemotherapeutic agent for patients with advanced ovarian cancer.

**Key words** Platinum analog · Nedaplatin · Chemotherapy · Ovarian cancer

## Introduction

The platinum compounds, cisplatin and its analog carboplatin, are important chemotherapeutic agents used in the initial treatment of ovarian and other solid cancers [10, 13, 15, 16]. Unfortunately, most patients who initially achieve a complete response to platinum-based therapy develop resistance to these drugs [13, 15]. Additionally, many patients who are treated with cisplatin develop dose-limiting neurotoxicity, hearing loss, and nephrotoxicity [3, 8]. Although carboplatin is more myelosuppressive than the parent agent, cisplatin, it is associated with significantly less renal, neurological and ototoxicity; however, it demonstrates almost complete crossresistance to cisplatin [4, 12]. Thus, research is ongoing to further reduce toxicities, and circumvent resistance to standard platinum agents (e.g. through the use of chemomodulating agents), and also to develop noncrossresistant analogs.

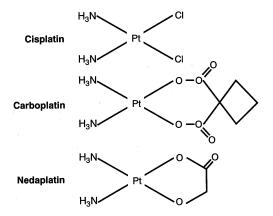


Fig. 1 Chemical structures of cisplatin, carboplatin and nedaplatin

Nedaplatin is an experimental platinum analog which has undergone recent clinical evaluation. It was selected from a series of platinum analogs for clinical evaluation based on its pronounced preclinical antitumor activity against solid tumors, virtual lack of nephrotoxicity and relatively low rate of neurotoxicity [20–22]. Results of phase I and phase II studies have revealed a spectrum of solid tumor activity similar to that of cisplatin [2, 5, 9, 14, 18, 19]. Its dose-limiting toxicity appears to be myelosuppression, particularly thrombocytopenia. Nedaplatin also has been associated with ototoxicity similar to that observed with cisplatin [2, 5].

The chemical structures of nedaplatin, cisplatin and carboplatin are displayed in Fig. 1. All three agents undergo hydrolysis, although at different rates, to form active divalent species which ultimately form DNA intrastrand adducts.

In this study, we performed a comparative examination of the in vitro activity of nedaplatin, cisplatin and carboplatin against fresh human ovarian tumor samples using the Hamburger-Salmon human tumor colony-forming assay (HTCA) [7, 11, 17].

### **Materials and methods**

## Tumor samples

Solid tumor biopsy specimens were obtained from 36 previously untreated patients with epithelial, ovarian cancer (FIGO stage III-IV) during the initial exploratory laparotomy and tumor debulking surgery. Specimens were transferred aseptically to our HTCA laboratory within 24 h of surgery. Solid tumor specimens were transported in medium (McCoys 5A, Irvine, Santa Ana, Calif.) supplemented with 10% fetal bovine serum (FBS), penicillin (100 units/ml), streptomycin (100 mg/ml) and L-glutamine (2 mM) (Irvine).

## Drug preparation

Tumor specimens underwent drug sensitivity testing with multiple standard and investigational agents. Cisplatin (clinical sample, Bristol-Myers Squibb Oncology, Princeton, N.J.) was reconstituted in distilled water to give 1 mg/ml cisplatin in 10 mg/ml mannitol. Further dilutions were made with 0.9% NaCl. Carboplatin (clinical sample, Bristol-

Myers Squibb Oncology) and nedaplatin (research sample, Shionogi and Company Ltd., Osaka, Japan) were reconstituted in distilled water.

#### HTCA method

The HTCA was performed with minor modification as previously described for epithelial cancers of the ovary [7, 11, 17]. Solid tumor samples were minced into 2–3-mm pieces and then further digested in 8 ml 0.15% type I collagenase, 0.015% DNase mixture (Sigma, St. Louis, Mo.) and 8 ml hypo-osmotic medium in a stirring flask for 1.0 h at 37 °C. The digest was then filtered through 8X sterile gauze and washed with McCoy's 5A medium supplemented with 10% FBS. The resulting cell suspension was examined and, if necessary, a 25- or 30-μm mesh filtration was used to obtain a single-cell suspension.

Drug incubations were performed in triplicate at  $37\,^{\circ}\text{C}$  for 1 h in McCoy's 5A medium supplemented with 10% FBS. Standard drug exposures were 0, 10, and 100 µg/ml for nedaplatin and cisplatin and 0, 100, and 1000 µg/ml for carboplatin.

After drug incubation, the tumor cells were washed twice with 5 ml McCoy's 5A medium supplemented with 10% FBS and plated as previously described [11, 17] Tumor cells were plated at a concentration of approximately 250 000 cells/plate. Plates were incubated for 14 to 21 days at 37 °C in an atmosphere containing 5% CO<sub>2</sub>, stained with INT stain (Pfaltz and Bauer, Stamford, Ct.) and colonies were counted on an Omnicon Image Analyzer [17]. The plant lectin, abrin (Sigma, St. Louis, Mo.), was used as a positive control in each experiment.

Estimation of percent tumor colony-forming unit (TCFU) survival at clinically achievable drug concentrations

The minimum platinum drug doses used in this study  $(10-100~\mu g/ml$  for 1 h) were rough approximations of clinically achievable drug concentrations as defined by the mean ultrafiltrable platinum plasma AUC following administration of standard doses for single agent chemotherapy of gynecologic cancers  $(16~\mu g\times h/ml$  for nedaplatin,  $9.8~\mu g\times h/ml$  for cisplatin, and  $80~\mu g\times h/ml$  for carboplatin) [6, 18, 23]. We refined our estimation of clinical activity further by predicting the percentage survival relative to control of individual tumor samples at a dose equal to the mean ultrafiltrable platinum plasma AUC using a least squares fitted model with log(dose +1) as the predictor.

## Results

## Ovarian tumor samples

A total of 36 viable tumor samples from chemotherapynaive patients with epithelial ovarian cancer with stage III and IV were tested against each of the three platinum analogs, using the HTCA.

In vitro drug sensitivity data

Shown in Table 1 are the in vitro chemosensitivity data (expressed as percent survival of TCFU growth as compared with control plates) for each of the 36 different fresh ovarian cancers exposed separately to each of the three platinum-containing analogs for 1 h. All three agents exhibited strong dose-response relationships against the majority of the 36 cancers. With chemosensitivity being defined as ≤50% survival of TCFUs as compared with control plates, 15 of 36 cancers (42%) were sensitive to

Table 1 Percent survival of TCFUs obtained from fresh human ovarian cancers following a 1-h exposure to three platinum analogs at the specified in vitro concentrations (NA data not available)

Table 2 IC<sub>50</sub> values (drug concentration associated with 50% inhibition of TCFUs relative to control) of platinum-containing analogs against 36 fresh human ovarian tumors (UNACH unachievable)

			<u> </u>				
Tumor sample number	Nedaplati 10 µg/ml	n Cisplatin 10 μg/ml	Carboplatin 100 µg/ml	Tumor sample numer	Nedaplatin (μg/ml)	Cisplatin (µg/ml)	Carboplatin (µg/ml)
8332	131	99	111	8332	UNACH	61	UNACH
8324	14	33	26	8324	5	10	49
8320	84	86	96	8320	UNACH	45	UNACH
8318	35	29	15	8318	6	8	25
8316	50	43	41	8316	29	11	97
8305	84	50	50	8305	72	20	100
8302	51	60	54	8302	12	13	68
8299	47	37	79	8299	14	14	1225
8297	54	68	61	8297	25	21	915
8295	68	34	65	8295	51	12	101
8287	22	9	15	8287	5	6	23
8285	91	84	102	8285	UNACH	53	UNACH
8260	95	87	82	8260	103	107	298
8256	18	36	38	8256	6	9	47
8249	94	55	138	8249	77	12	UNACH
8246	72	73	43	8246	50	UNACH	130
8228	28	21	18	8228	14	8	32
8206	94	23	42	8206	85	7	42
8204	45	58	85	8204	35	16	290
8202	29	41	39	8202	5	9	39
8198	39	30	96	8198	9	7	307
8192	90	44	62	8192	72	10	118
8183	46	74	75	8183	29	100	UNACH
8180	46	62	81	8180	11	12	167
8178	64	29	53	8178	12	7	50
8170	45	76	45	8170	8	19	59
8165	76	51	70			19	
			50	8165	28 33		124
8163	46	97		8163		110	177
8154	98	71	NA	8154	UNACH	UNACH	UNACH
8148	72	37 52	54	8148	28	9	62
8146	106	52	83	8146	52	10	357
8140	92	62	67	8140	112	23	556
8132	81	82	60	8132	UNACH	UNACH	UNACH
8127	60	26	73	8127	11	7	98
8120	19	8	45	8120	4	5	39
8118	58	41	38	8118	16	6	42
Number tumors tested	36	36	35	Number achievable/number			
Median % survival	59	50.5	60	tested	31/36	33/36	29/36
Number sensitive (%) <sup>a</sup>	15/36 (42)	18/36 (50)	14/35 (40)	Median IC <sub>50</sub> of all values	28.5	12	121
a Sensitivity is defined as ≤50% survival of TCFUs relative to control				Mean of achievable values ± standard error	$33\pm6$	24±5	194±51

plates

nedaplatin at a concentration of 10 µg/ml, 18 of 36 (50%) to cisplatin at 10 µg/ml and 14 of 35 (40%) to carboplatin at 100 μg/ml. The 100 μg/ml concentration of carboplatin is roughly comparable to the 10 µg/ml cisplatin concentration, based on carboplatin's larger clinically achievable AUC, slow in vitro activation rate, and prior in vitro HTCA training set analysis against human tumor cell lines. The chemosensitivity rates for the three drugs were not significantly different from one another.

The concentrations of each platinum analog associated with 50% survival of TCFUs (i.e. IC<sub>50</sub> values) are listed in Table 2. The median IC<sub>50</sub> values for nedaplatin, cisplatin and carboplatin were 28.5, 12 and 121 µg/ml, respectively. The corresponding mean (±standard error) IC<sub>50</sub> values were  $33\pm6$ ,  $24\pm5$ , and  $194\pm51$  µg/ml, respectively. There were no statistically significant differences between the IC<sub>50</sub> values of cisplatin and nedaplatin (P > 0.1, McNe-

mar's test). Carboplatin's IC50 values were artifactually high owing to its slow in vitro activation.

Based on the raw percent TCFU survival data presented in Table 1, we evaluated the cross-sensitivity and crossresistance of carboplatin and nedaplatin, relative to the activity of cisplatin. As shown in Table 3, of the 18 ovarian cancers sensitive to cisplatin (i.e. ≤50% survival of TCFUs), 10 (56%) also were sensitive to nedaplatin and 11 (61%) to carboplatin. Of the 18 cancers resistant to cisplatin (i.e. >50% survival of TCFUs), only 5 of 18 (28%) and 3 of 17 (18%) were sensitive to nedaplatin and carboplatin, respectively.

Human pharmacokinetic studies have shown that following the administration of clinically tolerable and biologically active doses of nedaplatin, cisplatin and carboplatin (i.e. 100, 100 and 400 mg/m<sup>2</sup>, respectively), mean plasma ultrafiltrable platinum AUCs are 16, 9.8 and 80 µg×h/ml,

**Table 3** Chemosensitivity (defined as  $\leq$ 50% of TCFUs relative to control at doses of 10 µg/ml for cisplatin and nedaplatin and 100 µg/ml for carboplatin) of fresh ovarian tumors to nedaplatin and carboplatin, relative to cisplatin

	Sensitive to nedaplatin 15/36 (42%)	Sensitive to carboplatin 14/35 (40%)
Sensitive to cisplatin 18/35 (50%)	10/18 (56%)	11/18 (61%)
Resistant to cisplatin 18/36 (50%)	5/18 (28%)	3/17 (18%)

respectively [6, 18, 23]. The estimated percent TCFU survivals of all the ovarian cancers included in this study at the clinically achievable plasma levels (i.e. mean plasma AUCs) of the three platinum analogs are shown in Table 4. Based on these calculations, the activities of nedaplatin, cisplatin and carboplatin were roughly comparable with 42% of tumors sensitive to nedaplatin and 36% sensitive to cisplatin and carboplatin at clinically achievable concentrations.

#### **Discussion**

The platinum compounds, cisplatin and carboplatin, continue as two of the most commonly used anticancer drugs in the treatment of a variety of solid tumors, including cervix, head and neck, lung and ovarian cancers [15]. Unfortunately, both inherent and acquired tumor resistance to these drugs limit their clinical utility [13]. Thus, there has been a continued search for more active and noncrossresistant platinum analogs. Nedaplatin is one such drug which has undergone clinical testing in Japan.

We compared the in vitro cytotoxicities of nedaplatin, cisplatin and carboplatin against 36 fresh ovarian cancers from chemotherapy-naive patients using the HTCA. Nedaplatin, cisplatin and carboplatin were tested at 10 and 100  $\mu$ g/ml dose levels and carboplatin was tested at 100 and 1000  $\mu$ g/ml. At the lowest drug concentration tested (i.e. 10  $\mu$ g/ml for nedaplatin and cisplatin and 100  $\mu$ g/ml for carboplatin) the percentage of tumor samples which were sensitive (as defined by 50% or less survival of TCFUs as compared with controls) was 42%, 50% and 40% for nedaplatin, cisplatin and carboplatin, respectively.

We have established previously that the clinical relevance of in vitro data may be increased by consideration of human pharmacokinetic data, especially the mean plasma concentration time product (AUC) during the dose selection process [1]. Human pharmacokinetic studies have shown that following the administration of clinically tolerable and biologically active doses of nedaplatin, cisplatin and carboplatin (i.e. 100, 100 and 400 mg/m², respectively), mean plasma ultrafiltrable platinum AUCs are 9.8, 16 and  $80~\mu g \times h/ml$ , respectively [6, 18, 23]. The minimum platinum drug doses used in this study were within the region of clinically achievable levels. We refined the estimation of

**Table 4** Estimated percent survival of TCFUs at clinically achievable drug levels (i.e. mean free platinum plasma AUC following administration of standard doses<sup>a</sup>)

Tumor sample number	Nedaplatir	n Cisplatin	Carboplatin
8332	105	73	113
8324	31	50	44
8320	107	69	104
8318	33	46	33
8316	59	52	52
8305	68	61	53
8302	45	55	48
8299	47	55	70
8297	57	62	68
8295	65	53	53
8287	29	39	31
8285	73	71	82
8260	73	76	63
8256	31	48	43
8249	68	53	88
8246	64	78	55
8228	48	45	37
8206	69	42	42
8204	58	58	62
8202	32	47	41
8198	39	45	62
8192	67	51	54
8183	58	74	74
8180	43	53	58
8178	45	44	44
8170	40	61	46
8165	59	51	55
8163	60	78	57
8154	94	79	85
8148	58	49	47
8146	66	51	63
8140	71	62	65
8132	95	90	72
8127	43	43	53
8120	25	37	40
8118	50	42	42
Median % survival	58	53	55
Number sensitive (%) <sup>b</sup>	15/36 (42)	13/36 (36)	13/36 (36)

 $<sup>^</sup>a$  Mean free platinum AUCs were 16 µg×h/ml following a nedaplatin dose of 100 mg/m², 9.8 µg×h/ml following a cisplatin dose of 100 mg/m², and 80 µg×h/ml following a carboplatin dose of 400 mg/m²

clinical activity further by using a least squares fitted linear model to estimate the percent survival of TCFUs at the mean plasma AUCs listed above for the three platinum agents. The estimated percentage of sensitive tumors at the clinically achievable AUC was 42% for nedaplatin and 36% for cisplatin and carboplatin. Thus, nedaplatin appears to have comparable activity to cisplatin and carboplatin on the basis of the degree of sensitivity at the minimum dose tested, as well as at the clinically achievable AUC.

We showed that nedaplatin, as well as carboplatin, appear relatively crossresistant with cisplatin in vitro. For example, only 28% of the ovarian cancers resistant to cisplatin were sensitive to nedaplatin and only 18% were sensitive to carboplatin. Other studies have shown a partial crossresistance of nedaplatin to other platinum agents [2, 9]. Thus, nedaplatin, like carboplatin, may not prove useful as

b Sensitivity defined as ≤50% survival of TCFUs compared to control

a salvage agent in patients with cisplatin-refractory (or carboplatin-refractory) tumors; however, because of its relative lack of associated neurotoxicity and nephrotoxicity and high bioavailability, nedaplatin merits further clinical trials in both previously untreated and treated patients with advanced ovarian cancers.

#### References

- Alberts DS, Salmon SE, Chen HSG, Moon TE, Young L, Surwit EA (1981) Pharmacologic studies of anticancer drugs with the human tumor stem cell assay. Cancer Chemother Pharmacol 6:253
- Ariyoshi Y, Ota K, Wakui A, Majima H, Nittani H, Ogawa M, Inuyama Y, Yoshida O, Taguchi T, Kimura I, Kato T (1988) Phase I study of (glycolato-0-0') diammineplatinum(II) (254-S). Proc Am Soc Clin Oncol 7:222
- Calvert AH, Harland SJ, Newell DR, Siddik ZH, Jones AC, McElwain TJ, Raju S, Wiltshaw E, Smith IE, Baker JM, Peckham MJ, Harrap KR (1982) Early clinical studies with cisdiamminedichloroplatinum(II) (cisplatin). Cancer Chemother Pharmacol 9:140
- Evans BD, Raju KS, Calvert AH, Harland SJ, Wiltshaw E (1983) Phase II study of JM8, a new platinum analog in advanced ovarian cancer. Cancer Treat Rep 67:997
- Fukuda M, Shinkai T, Eguchi K, Sasaki Y, Tamura T, Ohe Y, Kojima A, Oshita F, Hara K, Saijo N (1990) Phase II study of (glycolato-0-0') diammineplatinum(II), a novel platinum complex, in the treatment with non-small-cell lung cancer. Cancer Chemother Pharmacol 26:393
- Goel R, Andrews PA, Pfeifle CE, Abramson S, Kirmani S, Howell SB (1990) Comparison of the pharmacokinetics of ultrafilterable cisplatin species detectable by derivatization of diethyldithiocarbamate with atomic absorption spectroscopy. Eur J Cancer 26:21
- Hamburger AW, Salmon SE, Kim MB, Trent JM, Soehnlen BJ, Alberts DS (1978) Direct cloning of human ovarian carcinoma cells in agar. Cancer Res 38:3438
- 8. Hamers FPT, Gispen WH, Neijt JP (1991) Neurotoxic side-effects of cisplatin. Eur J Cancer 27:372
- Inuyama Y, Miyake H, Horiuchi M, Hayasaki K, Komiyama S, Ota K (1992) An early phase II clinical study of cis-diammine glycolatoplatinum, 254-S, for head and neck cancers. Jpn J Cancer Chemother 19:863
- Kelland LR (1993) New platinum antitumor complexes. Crit Rev Oncol Hematol 15:191

- Leibovitz A, Liu R, Hayes C, Salmon SE (1983) A hypo-osmotic medium to disaggregate tumor cell clumps into viable and clonogenic single cells for the human tumor stem cell clonogenic assay. Int J Cell Cloning 1:478
- Mangioni C, Bolis G, Pecorelli S, Bragman K, Epis A, Favalli G, Gambino A, Landoni F, Presti M, Terri W (1990) Randomized trial in advanced ovarian cancer comparing cisplatin and carboplatin. J Natl Cancer Inst 81:434
- 13. Muggia FM, Los G (1993) Platinum resistance: laboratory findings and clinical implications. Stem Cells 11:182
- 14. Ota K, Wakui A, Majima H, Niitani H, Inuyama Y, Ogawa M, Ariyoshi Y, Yoshida O, Taguchi T, Kimura I (1992) Phase I study of a new platinum complex 254-S, cis-diammine (glycolato)-platinum(II). Jpn J Cancer Chemother 19:855
- Ozols RF, Young RC (1984) Chemotherapy of ovarian cancer. Semin Oncol 11:251
- Rosenberg B (1985) Fundamental studies with cisplatin. Cancer 55:2303
- Salmon SE, Hamburger AW, Soehnlen B, Durie BG, Alberts DS, Moon TE (1978) Quantitation of differential sensitivity of humantumor stem cells to anti-cancer drugs. N Engl J Med 298:1321
- Sasaki Y, Tamura T, Eguchji K, Shinkai T, Fujiwara Y, Fukuda M, Ohe Y, Bungo M, Horichi N, Niimi S (1989) Pharmacokinetics of (glycolato-0,0')-diammine platinum (II), a new platinum derivative in comparison with cisplatin and carboplatin. Cancer Chemother Pharmacol 23:243
- Sasaki Y, Amano T, Morita M, Shinkai T, Eguchi K, Tamara T, Ohe Y, Kojima A, Saijo N (1991) Phase I study and pharmacological analysis of cis-diammine (glycolato)platinum (254-S; NSC 375101D) administered by 5-day continuous intravenous infusion. Cancer Res 51:1472
- Sasaki Y, Shinkai T, Eguchi K, Tamura T, Ohe Y, Ohmori T, Saijo N (1991) Prediction of the antitumor activity of new platinum analogs based on their ex vivo pharmacodynamics as determined by bioassay. Cancer Chemother Pharmacol 27:263
- Shiratori O, Kasai H, Uchida N, Takeda Y, Totani T, Sato K (1985) Antitumor activity of 254-S, a platinum complex, in rodents. In: Ishigami J (ed) Recent advances in chemotherapy. University of Tokyo Press, Tokyo, p 635
- 22. Totani T, Aono K, Adachi Y, Komura M, Shiratori O, Sato K (1988) Bidentate hydroxycarboxylic acid platinum(II) complexes with antitumor activity. In: Marino N (ed) Platinum and other metal coordination compounds in cancer chemotherapy. Martinus Nijhoff, Boston, p 744
- Warmerdam WC van, Rodenhuis S, Telligen O van, Maes RAA, Beijen JH (1994) Validation of a limited sampling model for carboplatin in a high-dose chemotherapy combination. Cancer Chemother Pharmacol 35:179